
Publisher's PDF, also known as Version of record

License (if available):
CC BY-NC

Link to published version (if available):
10.1093/eurheartj/ehz459

Link to publication record in Explore Bristol Research

PDF-document

This is the final published version of the article (version of record). It first appeared online via OUP at https://doi.org/10.1093/eurheartj/ehz459. Please refer to any applicable terms of use of the publisher.

University of Bristol - Explore Bristol Research

General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available: http://www.bristol.ac.uk/pure/about/ebr-terms
Single systemic transfer of a human gene associated with exceptional longevity halts the progression of atherosclerosis and inflammation in ApoE knockout mice through a CXCR4-mediated mechanism

Annibale Alessandro Puca1,2*, Albino Carrizzo3, Chiara Spinelli1, Antonio Damato3, Mariateresa Ambrosio3, Francesco Villa1, Anna Ferrario1, Anna Maciag1, Francesco Fornai3,4, Paola Lenzi4, Valentina Valenti5, Flavio di Nonno3, Giulio Accarino2, Michele Madonna3, Maurizio Forte3, Gaetano Cali6, Andrea Baragetti7, Giuseppe Danilo Norata7,8, Alberico Luigi Catapano7,9, Monica Cattaneo1, Raffaele Izzo10, Valentina Trimarco10, Francesco Montella10, Francesco Versaci5,11, Alberto Auricchio12,13, Giacomo Frati14, Sebastiano Sciarretta3,14, Paolo Madeddu15, Elena Ciaglia2, and Carmine Vecchione2,3*,†

1Ageing Unit, IRCCS MultiMedica, Via G. Fantoli 16/15, 20138 Milan, Italy; 2Department of Medicine, Surgery and Dentistry, “Scuola Medica Salernitana” University of Salerno, Via S. Allende, 84081 Baronia (SA), Italy; 3IRCCS Neuromed, Loc. Camerelle, 86077 Pozzilli (IS), Italy; 4Department of Translational Research and New Technologies in Medicine and Surgery, University of Pisa, via Roma 55, 56126 Pisa, Italy; 5UOC Cardiologia Ospedale Santa Maria Girent, 00100 Latina, Italy; 6Department of Endocrinology and Experimental Oncology Institute, CNR, Via Sergio Pansini, 80131 Naples, Italy; 7Department of Pharmacological and Biomolecular Sciences, Università degli Studi di Milano, via Vanvitelli 32, 20129 Milan, Italy; 8Società Italiana per lo Studio della Ateriosclerosi (SISA) Centro Aterosclerosi, Bassini Hospital, Cinisello Balsamo, 20092 Milan, Italy; 9IRCCS Multimedica Hospital, 20099 Sesto San Giovanni Milan, Italy; 10Department of Advanced Biomedical Sciences, University Federico II of Naples, 80131 Naples, Italy; 11Department of Cardiovascular Disease, Tor Vergata University of Rome, 00133 Rome, Italy; 12Telethon Institute of Genetics and Medicine (TIGEM), 80078 Pozzuoli (Na), Italy; 13Department of Advanced Biomedical Sciences, Federico II University, 80131 Naples, Italy; 14Department of Medical and Surgical Sciences and Biotechnologies, Sapienza University of Rome, via Faggiana, 40100 Latina, Italy; and 15Bristol Medical School (Translational Health Sciences), Bristol Heart Institute, University of Bristol, Upper Maudlin Street, Bristol BS2 8HW, UK

Received 3 January 2019; revised 13 April 2019; editorial decision 6 June 2019; accepted 22 June 2019

Aims
Here, we aimed to determine the therapeutic effect of longevity-associated variant (LAV)-BPIFB4 gene therapy on atherosclerosis.

Methods and results
ApoE knockout mice (ApoE-/-) fed a high-fat diet were randomly allocated to receive LAV-BPIFB4, wild-type (WT)-BPIFB4, or empty vector via adeno-associated viral vector injection. The primary endpoints of the study were to assess (i) vascular reactivity and (ii) atherosclerotic disease severity, by Echo-Doppler imaging, histology and ultrastructural analysis. Moreover, we assessed the capacity of the LAV-BPIFB4 protein to shift monocyte-derived macrophages of atherosclerotic mice and patients towards an anti-inflammatory phenotype. LAV-BPIFB4 gene therapy rescued endothelial function of mesenteric and femoral arteries from ApoE-/- mice; this effect was blunted by AMD3100, a CXC chemokine receptor type 4 (CXCR4) inhibitor. LAV-BPIFB4-treated mice showed a CXCR4-mediated shift in the balance between Ly6Chigh/Ly6Clow monocytes and M2/M1 macrophages, along with decreased...
Introduction

Atherosclerosis, a multi-factorial disease influenced by genetic and environmental factors, represents one of the leading causes of death in industrialized society. As it is a slowly progressing disease, it is vital to find new treatments able to halt plaque progression and the underlying inflammatory substrate. In this regard, chemokines and small chemotactic peptides represent a potential therapeutic target.

Recent evidence indicates that the chemokine stromal-cell derived factor-1 (SDF-1, also known as CXCL12) plays an important role in the angiogenesis process by binding to the C-X-C chemokine receptor type 4 (CXCR4) present in proangiogenic cells and leading them to sites of reparative processes. Here, the CXCR4 identifies transitional bone marrow precursors that replenish the mature monocyte pool for peripheral responses. Here, the CXCR4 acts as a chemokine receptor on proangiogenic cells and leading them to sites of reparative processes by binding to the C-X-C chemokine receptor type 4 (CXCR4) on proangiogenic cells and leading them to sites of tissue damage. Among myeloid cells, the high expression of CXCR4 identifies transitional bone marrow precursors that replenish the mature monocyte pool for peripheral responses. Here, the recruitment of Ly6Clow monocytes has resulted essential in plaque regression through their proper differentiation into M2 macrophages. However, the precise role of CXCR4/CXCL12 in atherosclerosis remains controversial. Pharmacological disruption of CXCR4 significantly aggravates diet-induced atherosclerotic lesion development in ApoE−/− mice. In addition, a common allele variant of the CXCR4 locus has been associated with atherosclerotic coronary heart disease. However, it has been also reported that high expression of CXCR4 in mouse heart increases infarct size, reduces cardiac function and leads to recruitment of inflammatory cells. Furthermore, expression of CXCR4 and CXCL12 is reportedly increased in Ly6Cmonocyes and macrophages within both stable and unstable carotid atherosclerotic plaques compared with healthy vessels, both at mRNA and protein levels.

Long-living individuals (LLIs) delay or escape atherosclerosis-related cardiovascular disease (CVD). We have previously found that LLIs are enriched for a longevity-associated variant (LAV) in BPIFB4, a gene that encodes a secreted protein, designated BPIFB4, which has been implicated in atherosclerotic disease. This LAV of BPIFB4 reduces the atherogenic process and skews macrophages towards an M2-resolving phenotype through modulation of CXCR4, thus opening up novel therapeutic possibilities in cardiovascular disease.

Translational perspective

As the main risk factor for cardiovascular diseases (CVDs) is progressive ageing of the population, unravelling the secrets of healthy ageing may be the only way to limit CVD disabilities. Here, we show that the favourable phenotype of long-living individuals can be transferred by gene therapy with the longevity-associated variant (LAV) of BPIFB4 to animal models. Indeed, the ability of LAV-BPIFB4 to finely tune endothelial function and pro/anti-inflammatory balance, as well as the fine correlation between a high BPIFB4 plasma level and a blunted atherogenic process in the clinical setting, makes LAV-BPIFB4 a suitable candidate tool for the treatment of atherosclerosis and its related CVD complications.

Conclusion

Transfer of the LAV of BPIFB4 reduces the atherogenic process and skews macrophages towards an M2-resolving phenotype through modulation of CXCR4, thus opening up novel therapeutic possibilities in cardiovascular disease.

Keywords

Atherosclerosis • Low-density lipoprotein • Vascular function • Immune system
Methods

Animal models
All animal studies were performed in accordance with approved protocols by the IRCCS Neuromed Animal Care Review Board and by the Istituto Superiore di Sanità, Rome (number: 1163/2015-PR) and were conducted according to EU Directive 2010/63/EU for animal experiments. Animal models’ details and treatment are reported in Supplementary material online.

Cloning, vector production and in vivo gene therapy
Ten-week-old ApoE−/− male mice were treated with 1 x 10¹³ GC/kg of AAV-GFP, AAV-WT-BPIFB4, or AAV-LAV-BPIFB4, as described in the Supplementary material online.

Echo-Doppler analysis
Detailed procedure is available in the Supplementary material online.

Histology and lesion analysis
Detailed procedure is available in the Supplementary material online.

Transmission electron microscopy and post-embedding immune-cytochemistry
Transmission electron microscopy method is available in the Supplementary material online.

Ex vivo transfection of mouse vessels
Detailed procedure is available in the Supplementary material online.

In vitro studies on human monocyte-derived macrophages
Detailed procedure is available in the Supplementary material online.

Cytokine detection
Cytokines were analysed by beads-based multiplex ELISA (LEGENDplex, Biolegend, USA) as described in the Supplementary material online.

Patient cohorts
The patient enrolled for the evaluation of carotid stenosis, intima-media thickness, BPIFB4 plasma levels, and genotype characterization are described in the Supplementary material online.

Ex vivo studies of human vessels
The study protocol was approved by the local ethics committees of IRCCS Neuromed and done in accordance with the Declaration of Helsinki. All participants gave written informed consent. Institutional review board approval was obtained from IRCCS Neuromed (No. 20160106-1006). A detailed procedure is reported in the Supplementary material online.

Western blotting
Western blotting was performed on pooled protein extracts from mice vessels or on total segments of human vessels, as reported in the Supplementary material online.

Statistical analysis
Statistical analyses are available in the Supplementary material online.

Results

LAV-BPIFB4 reduces endothelial dysfunction
We examined the effects of LAV-BPIFB4 gene therapy in ApoE knockout mice, using GFP- and WT-BPIFB4-treated mice for comparison. Genes were delivered using adeno-associated viral (AAV) vectors injected into femoral arteries, with efficiency properly monitored in liver, myeloid cells, plasma, and vascular cells (Supplementary material online, Figures S1–S5 and Methods). In addition, each group was randomized to receive a CXCR4 antagonist, AMD3100, or vehicle to verify the effect of the receptor on the primary endpoints of the treatment (Figure 1A). The primary endpoint indicates no difference among groups with regard to KCl- or phenylephrine-induced vasoconstriction and endothelium-independent relaxation to nitroglycerine. Likewise, no difference was observed in the presence or absence of the CXCR4 inhibitor (Figure 1B–D and Supplementary material online, Figure S6A–D). In contrast, vessels obtained from ApoE knockout mice treated with AAV-LAV-BPIFB4 showed a complete rescue of acetylcholine-mediated endothelial vasorelaxation both in mesenteric and femoral arteries (Figure 1E and Supplementary material online, Figure S6D). Interestingly, AMD3100, a non-peptide antagonist of CXCR4, abolished the beneficial effect of AAV-LAV-BPIFB4 gene therapy (Figure 1E and Supplementary material online, Figure S6D).

Western blot analyses of mesenteric arteries explanted from AAV-LAV-BPIFB4-treated mice showed an increase of total and phosphorylated (serine 75) BPIFB4 protein (Figure 1F). Likewise, AAV-LAV-BPIFB4 increased the phosphorylation of eNOS at serine 1177 and of PKCa at threonine 497 (Figure 1F), in accordance with our previous published results.9 eNOS is required for CCL12/CXCR4-mediated endothelial actions and different PKC isoforms mediate CXCR4 phosphorylation/activation.13 These phosphorylation changes were blunted by AMD3100 treatment, thus confirming the importance of CXCR4 and upstream activators in LAV-BPIFB4-induced vasorelaxation (Figure 1F). Of note, the action of AAV-LAV-BPIFB4 was not dependent on total cholesterol or LDL circulating levels, which remained similarly elevated in all after AAV-LAV-BPIFB4 treatment (Supplementary material online, Table S1). Moreover, LAV-BPIFB4 exerted a protective effect on oxLDL-induced endothelial dysfunction (Supplementary material online, Figure S7).

LAV-BPIFB4 halts vascular plaque formation
We next considered the other primary endpoint of the in vivo study, namely the progression of atherosclerosis as assessed by Echo-Doppler and histology. Ultrasound scanning of aortic arch with epi-aortic vessels revealed the presence lipid plaques in ApoE knockout mice treated with AAV-GFP and AAV-WT-BPIFB4, whereas AAV-LAV-BPIFB4 gene therapy reduced the formation of vascular plaques (Figure 2A). The quantification of the abundance of lipid streaks in aorta, which represent initial structural changes detectable in atherosclerosis, confirmed that LAV-BPIFB4 reduced vascular damage as compared to all other groups (Figure 2B). Moreover, the effect of LAV-BPIFB4 was lost in AMD3100-treated mice (Figure 2A and B), thus corroborating the CXCR4-mediated protective role of LAV-BPIFB4 on the onset of the atherogenic process.
Ultrastructural evaluation of the aorta, femoral, and mesenteric arteries from ApoE knockout mice showed alterations in endothelial cells, such as cytosolic derangement, diluted cytoplasm, detachment of the endothelial membrane, and broken plasma membrane. Gene therapy with AAV-LAV-BPIFB4 preserved the regular architecture of the vascular endothelium, again through a CXCR4-dependent mechanism (Figure 2C–E).

An imbalance of the mononuclear phagocytic system within the vascular wall is pivotal in influencing plaque initiation and progression.14 Hence, we next verified the ability of LAV-BPIFB4 to interfere with the above mechanism by staining harvested aortic arch with CD68+14, a marker highly expressed by monocytes and lesional macrophages, and with α-smooth muscle actin and collagen, which represent important compositional features of the atherosclerotic lesions. Results indicated that LAV-BPIFB4 reduces macrophages infiltration without loss of smooth muscle cells, a finding suggesting not only a slowing of the atherogenic process but also plaque stabilization.15,16 The collagen composition in the aortic arch of LAV-treated mice, evaluated by Sirius red positive area, showed a thicker fibrous cap than in vessels from other treatments, indicating a role for LAV-BPIFB4 in containing the onset and progression of the atherogenic processes (Figure 2F). In agreement with these findings, femoral arteries stained with the mono-macrophage marker MOMA-2 showed a marked reduction of positive cells in vessels from AAV-LAV-BPIFB- infected mice when compared with controls, an effect partially abolished by co-treatment with AMD3100 (Supplementary material online, Figure S5).

Figure 1 Overexpression of LAV-BPIFB4 improves the vascular reactivity of ApoE null mice fed a high-fat diet, and a CXCR4 inhibitor abolishes this protective effect. (A) Experimental protocol; (B) vascular response of ex vivo mesenteric arteries from ApoE knockout mice to potassium (80 mmol/L KCl) and (C) the dose–responses to phenylephrine, (D) acetylcholine, and (E) nitroglycerine after 1 month of AAV-LAV-BPIFB4 treatment. AAV-GFP was used as a control. Values are mean ± standard deviation of eight independent experiments. B, C, D, E two-way ANOVA followed Tukey’s multiple comparisons test. Numbers next to the curve show adjusted P-values. (F) Representative western blot (left) and densitometric analysis (right) conducted on mesenteric artery lysates. Values are mean ± standard deviation (N = 3). One-way ANOVA followed Tukey’s multiple comparisons test. Numbers above square brackets show adjusted P-values.
LAV-BPIFB4 regulates the peripheral pool of monocytes in a CXCR4-dependent manner

The observed reductions of CD68$^+$ cells at the aortic arch level and of MOMA-2 positive cells in femoral arteries may reflect a redistribution of circulating monocytes in response to the treatment. Indeed, as shown by western blot analysis, peripheral monocytes and progenitor bone marrow myeloid cells expressed an enhanced level of BPIFB4 after gene therapy (Supplementary material online, Figure S5B). To test this, we next examined monocyte frequency in peripheral blood from mice treated with GFP, WT-BPIFB4, or LAV-BPIFB4. In particular, we focused on the two major subsets of murine monocytes: ‘classical’ Ly6C$^{high}$ and ‘non-classical’ Ly6C$^{low}$ cells. Flow cytometry analyses demonstrated LAV-BPIFB4 gene therapy causes a reduction of Ly6C$^{low}$ monocytes and an increase of Ly6C$^{high}$ monocytes when compared with controls (Figure 3A).

Monocyte polarization has been attributed to changes in the oscillation of CXCR4 expression on bone marrow precursor cells, which serve for the replenishment of the peripheral pool of Ly6C$^{high}$ monocytes. Additionally, a substantial portion of Ly6C$^{high}$ monocytes originates from the spleen. We confirmed that these central sites play a role in the observed peripheral polarization induced by gene therapy. In fact, LAV-BPIFB4, but not WT-BPIFB4 or GFP, increased the percentage of CXCR4$^+$ Ly6C$^{high}$ cells in the bone marrow and spleen of ApoE knockout mice. A similar trend was observed in peripheral
Importantly, AMD3100 contrasted the improved effect of LAV-BPIFB4 on Ly6Chigh cell frequency, thus supporting a role for CXCR4 in these phenomena (Figure 3C). Results of multiplex bead-based immunoassay showed that LAV-BPIFB4 gene therapy increases the peripheral blood levels of IL-23 and IL-27, effects contrasted by AMD3100 (Figure 3D and E).

**LAV-BPIFB4 induces an enrichment of M2 splenic macrophages and reduces the proliferative state of T cells**

IL-23 and IL-27 are mainly secreted by monocytes and macrophages, and they have been recently implicated in suppression of atherosclerosis, partly controlling myeloid cell accumulation. As Ly6C⁷ conversion to M2 macrophages has been seen to drive plaque regression in different models of atherosclerosis, we sought to examine whether LAV-BPIFB4 could shift the phenotype of macrophages towards the pre-resolving M2 (alternatively activated) state. Due to the limited macrophage yield from the vessel wall, we focused on readily available macrophages derived from the spleen. Flow cytometry analysis of CD206/CD86, a marker of the relative proportion of M2 vs. M1 cells, indicated that LAV-BPIFB4 increased M2 monocytes in the spleen of ApoE knockout mice (Figure 4A and B). LAV-BPIFB4-induced polarization of monocytes towards an M2 phenotype was prevented by concomitant CXCR4 inhibition (Figure 4B).

M2 macrophages can reduce pro-inflammatory cytokine secretion and dampen inflammatory responses. Therefore, we next compared the proliferative state of CD4+ and CD8+ T cells in the spleen and peripheral blood of ApoE knockout mice treated with LAV-BPIFB4, WT-BPIFB4, or GFP (Figure 4C and D). Both LAV-BPIFB4 and WT-BPIFB4 reduced the abundance of Ki-67+ CD3 T cells, with AMD3100 inhibiting this effect only in the LAV-BPIFB4 group in peripheral blood compartment (Figure 4D). Looking at CD3 subfractions mostly affected by these changes, we found that LAV-BPIFB4 specifically reduced the abundance of proliferating cytotoxic CD8+ T cells in the spleen and in a significant manner in peripheral blood (Figure 4D).
In vitro exposure of PB-MNCs to LAV-BPIFB4 protein induces M2 macrophage differentiation

In order to confirm a direct effect of LAV-BPIFB4 on macrophage polarization, we tested the effect of human recombinant LAV-BPIFB4 protein on CD14+ PB-MNCs collected from atherosclerotic patients, which were induced to differentiate ex vivo into M1 or M2 macrophages, using a CellXVivo™ Kit, or into MPl macrophages, after exposure to autologous plasma. After 72 h of cytokine priming, cells were maintained in culture for an additional 72 h in the presence of LAV-BPIFB4 recombinant protein or vehicle. As shown in Figure 5A, cells harvested at the end of the conditioning included M1 macrophages displaying the canonical CD14+/CD206−/CD163− phenotype, and M2 macrophages characterized by the CD14+/CD206+/CD163+ phenotype. Flow cytometry analyses confirmed the ability of MPl to induce an M1 phenotype, whereas LAV-BPIFB4 polarized macrophages away from the M1 phenotype and towards a classical M2 anti-inflammatory state (Figure 5A and B). Of note, co-treatment with AMD3100 partially blunted the acquisition of the M2 phenotype induced by LAV-BPIFB4. This confirms the direct involvement of CXCR4 in LAV-BPIFB4’s polarizing effects (Figure 5C).

Finally, in order to strengthen the concept that LAV-BPIFB4 treatment can modify the inflammatory milieu within the vasculature, we measured the levels of pro- and anti-inflammatory mediators secreted by organotypic cultures of human atherosclerotic vessels. As shown in Figure 5D, stimulation with LAV-BPIFB4 protein reduced the levels of pro-inflammatory IL-1β and TNF-α, while increasing atheroprotective IL-33,25,26. Moreover, LAV-BPIFB4 improved endothelial-mediated vasorelaxation and eNOS phosphorylation of human atherosclerotic vessels (Figure 5E).

Figure 4 Enhanced enrichment of M2 macrophages in the spleen of AAV-LAV-BPIFB4 ApoE knockout mice fed a high-fat diet. (A and B) CD45hiCD11b+ F4/80+ splenic macrophages were additionally stained with flow cytometric markers CD206+ or CD86+ to discern the CD11b+ F4/80+CD206 M2 type from CD11b+ F4/80+CD86 M1 type of splenic macrophages. A representative dot plot panel (left) is presented. The graph on the right reports the mean ± standard deviation of ratios of M2 vs. M1 splenic macrophages (CD206/CD86 ratio) from all recipient mice (N=3–5 per group). MFI stands for mean fluorescence intensity of selected markers. (C) Analysis of the proliferation of splenic and blood CD3+ T cells in ApoE−/− mice infected with AAV-LAV-BPIFB4, AAV-WT-BPIFB4 or AAV-empty vector, treated or not with AMD3100. Representative dot plot showing changes of percentage Ki-67 expression in both spleen and blood TCD3+ cells. (D) Bar graph reporting the percentage ± standard deviation of Ki67+, CD3+ gated T cells. Percentage of Ki-67+ expression in CD4+ and CD8+ T cell subsets from the spleen and the blood of recipient mice were determined. The bar graph reports the percentage ± standard deviation. Statistical analysis by two-way ANOVA with post hoc Fisher’s Least Significant Difference (LSD) test was conducted. Numbers above square brackets show unadjusted LSD P-values.
High plasma BPIFB4 associates with a reduction of atherosclerotic risk

Starting from a population of 2606 individuals fully characterized in the ‘Progressione della Lesione Intimale Carotidea’ (PLIC) study, we focused on two groups of subjects stratified for the presence of subclinical carotid atherosclerosis (respectively, lower or higher than 25% of carotid stenosis; N = 90) (Table 1), and evaluated plasma levels of BPIFB4. Interestingly, our data revealed that in patients with no
subclinical carotid atherosclerosis, the protein’s concentration was significantly higher as compared to patients with carotid stenosis (Figure 6A). The functional role of the circulating BPIFB4 was magnified when examining its association with IMT stratification of a cohort of 22 consecutively enrolled non-smokers, non-diabetics without previous CV events and not on statin therapy, belonging to the Campania Salute Network Registry, with hypertension as the singular cardiovascular risk factor and an absence of other comorbidities (Table 2). As shown in Figure 6B, BPIFB4 was significantly higher in plasma from patients with IMT <2 mm, confirming a protective role of high protein levels, a finding in line with what was previously observed in the PLIC study. Moreover, genotype stratification analysis revealed that LAV carriers significantly more frequently had IMT <2 mm cohort, and that in this cohort, LAV-BPIFB4 carriers had a higher level of circulating BPIFB4 when compared with no-carriers (Figure 6C and D).

**Discussion**

In recent years, different approaches have been developed to counteract the progression of vascular atherosclerosis, including cholesterol-level lowering and inflammation modulation. Owing to the large numbers of inflammatory molecular and cellular mediators, it is unlikely that blockade of a single cytokine will be therapeutically effective. We report here new exciting results on the pleiotropic activity of LAV-BPIFB4 on different mechanisms of the atherogenic process. In particular, we demonstrate the efficacy of LAV-BPIFB4 in contrasting endothelial dysfunction, plaque formation/progression, inflammatory cytokine release, macrophage polarization, and T cell activation, in a murine model of atherosclerosis. These benefits were not associated with changes in the lipid profile. In addition, we provide ex vivo and in vitro evidence that these beneficial actions may extend to human vasculature until to be inversely associated to subclinical index of atherosclerosis in selected patients. Mechanistically, the effects of LAV-BPIFB4 seem to be attributable to a CXCR4-dependent mechanism.

Investigation of CXCR4 and CXCL12 expression revealed that the ligand and receptor are both up-regulated in stable and unstable carotid atherosclerotic plaques, especially in macrophages, compared with healthy vessels, both at the mRNA and protein level. It is, however, unclear if these expression changes are pathogenic or compensatory. Results from a recent study using a genetic approach argue for the second possibility: in fact, in ApoE knockout mice fed a high-fat diet, vascular-specific deletion of CXCR4 resulted in larger atherosclerotic lesions compared with their relative control. In line with this finding, reconstitution of LDLr(-/-) mice with autologous bone marrow infected with lentivirus encoding SDF-1α antagonist or CXCR4 degron, which affect proteasomal degradation of CXCR4, led to progressive plaque development, intraplaque haemorrhage and disease progression. Moreover, CXCR4 knockdown augmented the adhesive capacity of neutrophils and the activation state of circulating neutrophils. An epidemiological study followed by functional validation with the CRISPR/Cas9 system confirmed that genetic variation of CXCR4 can confer different susceptibility to coronary artery disease. Altogether, these findings open new therapeutic perspectives for the treatment of atherosclerosis through the targeting of the CXCL12/CXCR4 signalling pathway. Our study provides new evidence that this could be indeed a valuable option. In a previous investigation, we showed that BPIFB4 and CXCR4 expression concordantly distinguished healthy LLIs and correlated with maintained MNC migration towards CXCL12. Here, we demonstrate that LAV-BPIFB4 gene therapy protects from atherosclerosis through a mechanism involving the CXCR4 receptor in vascular cells and MNCs.

Long-living individuals have the unique ability to age without experiencing major CVD. The present study strengthens the novel concept that the healthy phenotype of LLIs can be efficiently transferred to murine models and cultured human tissues by the delivery of either LAV-BPIFB4 or a recombinant protein. LAV-BPIFB4 gene therapy succeeded in the two primary endpoints, namely improving endothelial dysfunction and reducing adverse vascular effects in ApoE knockout mice fed a high-lipid diet. The benefit of LAV-BPIFB4 was also evidenced at the ultrastructural level in vessels: it prevented the cytosolic derangement, cytoplasmic dilution, and irregular plasma membrane observed in different vascular districts of control mice.

**Table I** Clinical characteristics of patients from the PLIC study

<table>
<thead>
<tr>
<th>Parameters</th>
<th>PLIC population (N = 90)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>73 (70–77)</td>
</tr>
<tr>
<td>Gender (n, women)</td>
<td>50</td>
</tr>
<tr>
<td>Smoking habit (n, active)</td>
<td>18</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>26.70 (23.90–29.60)</td>
</tr>
<tr>
<td>Waist (cm)</td>
<td>93.5 (87.0–99.2)</td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>130 (120–140)</td>
</tr>
<tr>
<td>DBP (mmHg)</td>
<td>80 (70–85)</td>
</tr>
<tr>
<td>Anti-hypertensives (n, yes)</td>
<td>46</td>
</tr>
<tr>
<td>Fasting glucose (mg/dL)</td>
<td>93.00 (86.00–104.25)</td>
</tr>
<tr>
<td>Oral lowering therapies (n, yes)</td>
<td>5</td>
</tr>
<tr>
<td>Total cholesterol (mg/dL)</td>
<td>202.06 (30.91)</td>
</tr>
<tr>
<td>HDL (mg/dL)</td>
<td>59.04 (12.42)</td>
</tr>
<tr>
<td>Triglycerides (mg/dL)</td>
<td>88.00 (70.75–116.25)</td>
</tr>
<tr>
<td>LDL (mg/dL)</td>
<td>123.40 (29.30)</td>
</tr>
<tr>
<td>ApoA-I (mg/dL)</td>
<td>151.64 (18.78)</td>
</tr>
<tr>
<td>ApoB (mg/dL)</td>
<td>105.93 (19.91)</td>
</tr>
<tr>
<td>Statins (n, yes)</td>
<td>0</td>
</tr>
<tr>
<td>ALT (UI/L)</td>
<td>16 (13–21)</td>
</tr>
<tr>
<td>AST (UI/L)</td>
<td>18 (16–22)</td>
</tr>
<tr>
<td>GGT (UI/L)</td>
<td>20 (14–27)</td>
</tr>
<tr>
<td>Creatinine (mg/dL)</td>
<td>0.85 (0.76–0.95)</td>
</tr>
<tr>
<td>GFR (Cockcroft–Gault formula, mL/min/1.73 m²)</td>
<td>69.35 (17.65)</td>
</tr>
<tr>
<td>Previous CVD (n, yes)</td>
<td>19</td>
</tr>
</tbody>
</table>

Data are reported as mean ± standard deviation for normally distributed variables (Shapiro–Wilk test) or median (interquartile range) for non-normally distributed variables (see Supplementary material online, Statistical Analysis section). ALT, alanine aminotransferase; ApoA-I, apolipoprotein A1; ApoB, apolipoprotein B; BMI, body mass index; CVD, cardiovascular disease; DBP, diastolic blood pressure; GGT, gamma-glutamyltranspeptidase; HDL, high-density lipoproteins; LDL, low-density lipoproteins; SBP, systolic blood pressure.
Interestingly, LAV-BPIFB4 gene therapy did not affect the serum cholesterol profile, but it did contrast the ability of oxidized cholesterol to induce endothelial dysfunction by positively modulating the inflammatory/immune background of atherosclerosis.32

In line with this, LAV-BPIFB4 redistributed the pool of monocyte subpopulations, redirecting them towards a pro-resolving phenotype. This was reflected by the increased abundance of CXCR4⁺Ly6Chigh monocytes in bone marrow and spleen, the two major tissue reservoirs of monocytes available to mobilize towards injured tissues. In the margination process, CXCR4 is considered the retention force in the vasculature. Therefore, we speculate that the higher level of CXCR4 in blood Ly6Chigh monocytes after LAV-BPIFB4 treatment in mice may finely tune the transit time into the circulation, completing a protective intravascular differentiation process. In this context, recent studies have provided new interesting insight into the regulatory mechanisms of monocytosis relevant to atherosclerosis.5,32 Accordingly, we documented an enrichment of M2 splenic macrophages, which can contribute to dampen T cell activation and proliferation in a CXCR4-dependent manner, as AMD3100 counteracted most of the LAV-BPIFB4-mediated actions on the mono-macrophage compartment. This latter result is in keeping with the reported ability of CXCR4 to promote the acquisition of the M2 phenotype in healthy monocyte-derived macrophages.33 Furthermore, the described influence of LAV-BPIFB4 on

**Figure 6** High plasma BPIFB4 detected in LAV-carrier patients is associated with reduced carotid stenosis and IMT < 2mm. Graphs showing correlation between: (A) BPIFB4 plasma level in patients with carotid stenosis >25% (N = 48) and in patients with no carotid atherosclerosis (N = 42), (B) BPIFB4 level in patients stratified for IMT <2 mm (N = 14) vs. IMT >2 mm (N = 8). (C and D) Correlation between BPIFB4 protein levels with intima media thickness (C) and with plasma concentration (D) after genotype stratification of WT- or LAV-carrier patients. Results are presented as mean ± standard deviation and were analysed by two-tailed non-parametric Mann–Whitney test. Numbers above lines show unadjusted P-values.

### Table 2  Characteristics of hypertensive patients from the Campania Salute Study

<table>
<thead>
<tr>
<th>Parameters</th>
<th>CS population (N = 22)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Clinical characteristics</strong></td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>69 ± 7</td>
</tr>
<tr>
<td>Sex (male/female)</td>
<td>11/14</td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>143.6 ± 15.7</td>
</tr>
<tr>
<td>DBP (mmHg)</td>
<td>89.56 ± 12.5</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>76.36 ± 11.9</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>163.1 ± 9.5</td>
</tr>
<tr>
<td>Cholesterol (mg/dL)</td>
<td>212.8 ± 47.4</td>
</tr>
<tr>
<td>HDL (mg/dL)</td>
<td>55.25 ± 10.9</td>
</tr>
<tr>
<td>TG (mg/dL)</td>
<td>130.5 ± 60.4</td>
</tr>
<tr>
<td>Glycaemia (mg/dL)</td>
<td>96.46 ± 12.1</td>
</tr>
<tr>
<td>Creatinine (mg/dL)</td>
<td>0.905 ± 0.19</td>
</tr>
<tr>
<td>Medication (%)</td>
<td></td>
</tr>
<tr>
<td>β-blocker</td>
<td>5 (20)</td>
</tr>
<tr>
<td>Ang-1l receptor antagonist</td>
<td>12 (48)</td>
</tr>
<tr>
<td>ACE-inhibitor</td>
<td>8 (32)</td>
</tr>
<tr>
<td>Diuretics</td>
<td>11 (44)</td>
</tr>
<tr>
<td>CCB</td>
<td>9 (36)</td>
</tr>
</tbody>
</table>

Data are reported as mean ± standard deviation. CCB, amlodipine/olmesartan medoxomil; DBP, diastolic blood pressure; HDL, high-density lipoproteins; SBP, diastolic blood pressure; TG, triglycerides.
the myeloid compartment is in agreement with the recorded increase in IL-23 and IL-27 in peripheral blood in the mouse model. Indeed, both cytokines modulate endothelial cell activation and myeloid cell recruitment at early and advanced stages of atherosclerosis, limiting the deleterious effects of the pathological events.34,35

Our study also provides compelling in vitro evidence that the LAV-BPIFB4 protein can exert similar beneficial effects in redirecting human monocyte-derived macrophages from an M1 pro-inflammatory to an M2 anti-inflammatory phenotype, as well as correcting the balance of cytokines, increasing eNOS phosphorylation, and improving endothelial function in human atherosclerotic vessels. All these beneficial events observed in the different anatomical districts might be finely orchestrated by the circulating levels of BPIFB4, which was found significantly increased in plasma of AAV-LAV-BPIFB4-treated mice after 1 month of gene therapy. Indeed, the undetectable level of GFP protein both in myeloid cells and aorta led us to speculate a bystander action of circulating BPIFB4 on the immune compartment and in vascular cells after its proper production and rapid secretion by transduced liver cells (Take home figure). Accordingly, elevated plasma concentration of the protein associated with protection against the onset of the atherosclerotic disease, regardless of risk factors, in two independent patient cohorts, a finding corroborating the protective role of the LAV-BPIFB4 isoform and strengthening its translational relevance.

Conclusion

In conclusion, the novel findings from this study highlight the potential of LAV-BPIFB4 in contrasting vascular and immune features of atherosclerosis. This supports an alternative therapeutic vision of the management of CVDs. Hence, we foresee that LAV-BPIFB4 therapy could be a viable option to delay the occurrence of age-related cardiovascular disease, replicating successful, healthy longevity in populations at risk.

Supplementary material

Supplementary material is available at European Heart Journal online.

Funding

This work was supported by: research funding from Cariplo Foundation (n.2016-0874) to AAP and CV; PRIN-20157ATSLF_009 to AAP and CV; Ministry of Health (RF-2016-02364864) to AAP and CV; E. Ciaglia was supported by a fellowship from Fondazione Umberto Veronesi (FUV 2017cod.1072, FUV 2018cod.2153,FUV 2019cod.2198).

Conflict of interest:

A.A.P. and C.V. own shares of LGV1 Inc. and have filed a patent. All other authors declare no financial or competing interests.

References


