The filamentous fungus *Byssochlamys fulva* strain IMI 40021 produces (±)-byssochlamic acid 1, its novel dihydroanalogue 2 and four related secondary metabolites. Agnestrides A, 17 and B, 18 constitute a novel class of seven-membered ring, maleic anhydride-containing (hence termed heptadride) natural products. The putative maleic anhydride precursor 5 for both nonadride and heptadride biosynthesis was isolated as a fermentation product for the first time and its structure confirmed by synthesis. Acid 5 undergoes facile decarboxylation to anhydride 6. The generic term *maleidrides* is proposed to encompass biosynthetically-related compounds containing maleic anhydride moieties fused to an alicyclic ring, varying in size and substituents.

The collective name “nonadrides” was introduced by Barton and Sutherland for fungal metabolites containing a nine-membered ring fused to one or two maleic anhydride moieties in 1965. The first of these, gluconic acid 3 and gluconic acid 4 were isolated from *Penicillium glaucum*, by Wijkman and reported in 1931. Soon afterwards the pseudo-symmetrical analogue, byssochlamic acid 1 was isolated from the common food contaminant *Byssochlamys fulva* by Raistrick and coworkers. However the structures of 1 and 4 were only established some 30 years later by X-ray analysis. Barton and coworkers subsequently reisolated 4 from *Penicillium purpurgenum* and began considering the biosynthesis of the nonadrides. The *cis* relative stereochemistry of the ethyl and propyl groups of byssochlamic acid was confirmed by X-ray analysis of the bis-p-bromophenylhydrazide. The absolute configuration of 1 was established by classical degradative methods, and subsequently confirmed by enantiospecific synthesis of (±)-byssochlamic acid.

All three metabolites, 1, 3 and 4, were hypothesised to be biosynthesised via alternative dimerisation modes of a C9/C10-maleic acid precursor 5 and/or 6, and evidence for this was obtained by feeding studies with radiolabelled 6. Many more nonadride metabolites have since been reported, e.g. heveadride 7, an isomer of 1, and others which are expected to be biosynthesised through dimerisation of differently substituted maleic anhydride units, including scytalidin 8, and castaneolide 9. More complex examples include rubratoxin A 10, and its dihydro-analogue rubratoxin B 11 and phomoidrides A 12 and B 13. Cornexistin 14, is a nonadride unique in having a single anhydride unit. Viburspiran 15 and zopfiellin 16, feature 8-membered carboyclic rings and so have been classed as octadrides. Notably, all of the compounds belonging to this family reported so far have been isolated from fungi.
of a new analogue, dihydro-byssochlamic acid 2; two novel metabolites agnestadride A 17 and agnestadride B 18, both containing 7 membered rings which we have therefore termed heptadrides; and as a natural product for the first time, the proposed nonadride precursor, maleic anhydride 5 along with its decarbonylated derivative 6.

*B. fulva*, was grown as described by Raistrick3 in static culture in Czapek-Dox liquid medium with glucose as the sole carbon source for between 7 and 30 days. The crude ethyl acetate extracts were analysed by LCMS. Byssochlamic acid 1 \((C_{18}H_{20}O_6)\) was detectable after 7 days of static fermentation (Fig. 1, 15.4 min) and large amounts \((>50 \text{ mg L}^{-1})\) could be isolated after 4 weeks. Its identity was confirmed as (+)-byssochlamic acid by comparison of 1D \(^1H\) and \(^{13}C\) NMR spectra, and optical rotation comparison with literature values.8 Detailed analysis of 2D NMR spectra (COSY, HSQC and HMBC) allowed all signals in the \(^1H\) and \(^{13}C\) NMR spectra to be fully assigned for the first time (Table S1, ESI†).

The second-most abundant component of the extract eluted at 11.3 min (Fig. 1). The new compound had a UV absorption characteristic for nonadrides \((\lambda_{max} 211, 260 \text{ nm})\), ionised well in the negative ESIMS mode and had the same fragmentation pattern by byssochlamic acid 1 – the only difference being that all of the peaks appeared at \(m/z\) values two units higher. HREIMS analysis confirmed the molecular formula to be \(C_{18}H_{20}O_6\) corresponding to a reduced form of 1. Dihydrobyssochlamic acid 2 was purified by mass-directed preparative HPLC. The NMR spectra (Table S1, ESI†) were compared with those of byssochlamic acid 1. A new peak at 95.9 ppm in the \(^{13}C\) NMR spectra correlated (HSQC) to H-10 at 5.71 ppm confirmed the presence of a hemiacetal in 2. Consistent with the loss of the carbonyl, the resonances of C-11 and C-9 appeared at higher chemical shifts (172.2 and 160.7 ppm respectively), and the C-8 resonance moved upfield to 130.6 ppm. Key HMBC signals included correlations between 1-CH\(_2\) protons (2.42/2.32 ppm) and C-8, C-9 and C-10; C-9, C-8 and C-11 but not C-10 correlated also to H-7 (2.81 ppm), consistent with the lactol hydroxyl being attached to C-10. Moreover, 6-CH\(_2\) (3.05/2.52 ppm) showed correlations with C-8, C-4, C-5 and C-13 (130.1, 139.1, 139.9 and 168.5 ppm respectively) but not C-11 or C-9 (171.8 and 160.3 ppm). Interestingly, closer inspection of the \(^1H\) NMR spectrum showed the presence of one major singlet at 5.71 ppm consistent with the hemiacetal adopting one major configuration (68%) but three other singlet methine signals were apparent at 5.80 (16%), 5.84 (5%), and 5.93 (11%) ppm respectively. While one of these minor components is likely to be due to the epimeric lactol, the other two signals may be due to regioisomeric lactols. Other nonadride lactol analogues are known, e.g. rubratoxin B 11\(^{13}\) and dihydro- and tetrahydroepiheveadride \((e.g. 7 \text{ } R = \text{ H, OH, } * = \text{ S})\) have been isolated from *Wicklowia aquatica*, along with the corresponding lactones.18 In phomoidride A 12, one of the anhydrides is found as a hemiacetal due to a transannular C-C bond, and in phomoidride B 13 as a full acetal due to further linkage to a side chain hydroxyl.14

LCMS chromatograms of extracts from older cultures of *B. fulva* (ca. 30 days) showed a new peak at 13.3 min with the same molecular weight as byssochlamic acid 1 but a slightly different UV spectrum. It was isolated by HPLC (~1 mg L\(^{-1}\) culture) and HREIMS confirmed it to have the same molecular formula \((C_{18}H_{20}O_6)\) as byssochlamic acid. However, \(^1H\) NMR analysis (‘Table S2, ESI†) showed the presence of a 1-butynyl side chain. The HSQC spectrum revealed only nineteen hydrogens attached to carbon, suggesting that the other belonged to a hydroxyl. The presence of a further propyl substituent linked to an otherwise uncoupled CH, and two mutually coupled methylenes were obvious from the COSY spectrum (Fig. 2a). The characteristic signals for one maleic anhydride were apparent at 143.3 (C-11), 165.3 (C-12), 164.8 (C-13) and 144.9 (C-14) ppm. The remaining 4 carbons resonances at 168.5 (C-6), 127.0 (C-5), 153.4 (C-8) and 104.0 (C-7) ppm were indicative of an \(\alpha,\beta\)-unsaturated ester/lactone and an acetal, consistent with the hydroxy lactone moiety indicted in bold in Fig. 2a. Further detailed analysis of the HMBC data (Fig. 2b) was consistent with the seven-membered carbocycle, which we name agnestadride A 17. The key correlations were seen from the butenyl H-4 (6.08 ppm) to C-5, C-6 and C-8, and from H-16 (3.54 ppm) to C-7, and 9-CH\(_2\) (3.01/2.66 ppm) to C-5, C-7 and C-8 to firmly locate the hydroxylactone ring. Similarly, correlations from 9-CH\(_2\) to C-10 (20.6 ppm) and C-11, and 10-CH\(_2\) (3.05/2.52 ppm) to C-11, C-12 and C-13 locate the maleic anhydride moiety.

A less polar new compound was detected (18.4 min in Fig. 1) and was isolated (1 mg L\(^{-1}\)) using mass-directed preparative HPLC. It had a mass of 314 (peak of \(m/z\) 313 \([M-H]^-\) in the negative ESIMS spectrum), corresponding to a dehydrated derivative of 17. The formula was confirmed by HREIMS analysis to be \(C_{18}H_{18}O_5\). In contrast with other colourless metabolites isolated from *B. fulva* compound 18 was bright yellow. The \(^1H\) NMR spectrum revealed

![Fig. 1](HPLC (DAD) chromatogram of *B. fulva* extract.)

![Fig. 2](a) COSY correlations of three isolated spin systems of 17. (b) HMBC correlations.)
that 18 also contained a pendant 1-butenyl side chain. A total of 18 protons were observed in the 1H NMR spectrum (Table S2, ESI†). There were three 13C NMR resonances (166.2, 164.6 and 163.1 ppm) characteristic for anhydride/unsaturated lactone carbonyls, and 6 quaternary olefinic carbons. The C-7 acetal and C-15 methine signals in 18 were absent, being replaced by two quaternary olefinic carbons. This data is fully consistent with compound 18, named agnestadride B, being a dehydrated form of agnestadride A. The structure is supported by HMBC correlations, from H-4 to C-5 and C-6, as well as from 17-CH3 to C-15. To our knowledge, no heptadrides have been previously reported as natural products. Baldwin and co-workers, investigating biomimetic dimerisation of putative nonadride maleic anhydride monomers isolated a heptadride structure 19 which was also reported as being an intense yellow compound.19

Barton and coworkers suggested that maleic anhydride 6 is the precursor of 1.6 Although indirect and direct evidence has accumulated for this hypothesis,6,20,21 anhydride 6 has never been observed as a component of B. fulva fermentations. Subsequent feeding studies on the phomoidrides utilising synthetic precursor analogues by Sulikowski and coworkers22 indicated that the monomeric nonadride precursor unit may be a carboxylated species, e.g. 5 for byssochlamic acid. In an attempt to find 5 or 6 in extracts of B. fulva, we searched extracted ion chromatograms of fresh extracts and identified a candidate peak at 7.7 min (Fig. 1). This metabolite formed ions of m/z 209.5, 165.5 in the negative ESIMS, which matched masses of ions expected for 5 ([M – H]− and [M – H – CO2]− respectively) and m/z 211.5, 193.3 in the positive ESIMS ([M + H]+ and [M + H – H2O]+ respectively). This compound was purified, but identification by full NMR (Table S3, ESI†) and HRESIMS analysis surprisingly showed it to be 6, the decarboxylated form of 5. Reinjection of 6 into the LCMS showed that it had a different retention time to 5, suggesting that it had decarboxylated during purification and/or subsequent analysis. So to confirm the structure and study its stability maleic anhydride 5 was synthesised as shown in Scheme 1 based on previous work by Sulikowski.23 Commercially available muconic acid 20 was protected as silyl ether 21 in 79% yield. (E)-2-(But-1-en-1-yl)benzo[d][1,3,2]dioxaborole 22 prepared via the hydrometallation of 1-butyne with catecholborane23 underwent Suzuki cross-coupling with 21 to give bromide 23 regio-selectively (as proven by HMBC). No bis-coupling was observed and the trans-geometry of the alkene was confirmed by a 16 Hz coupling constant in the 1H NMR spectrum. Palladium-catalysed coupling of the bromide 23 with silyl enol ether 24 (prepared separately by treatment of tert-butyl acetate with LDA and TBSCI)24 afforded ester 25 in an excellent yield of 93%. Removal of the TBS group with HF–pyridine was sluggish, giving yields of 26 typically between 30–46%. The resulting lactol 26 was then oxidised efficiently by Dess–Martin periodinane to 27 and final ester hydrolysis with TFA generated anhydride 5 in 66% yield.

The identity of naturally occurring 5 was confirmed by comparing its LCMS characteristics with the data collected for the synthetic product. The synthetic compound was observed to undergo decomposition to 6 spontaneously and decomposition was complete in under 48 h. The less polar peak at 20.6 min (Fig. S2, ESI†) had a similar UV spectrum to 5, however it did not ionize well (the ESI spectrum shows a m/z 165 with a m/z of 209 but only in very concentrated samples), which was consistent with the loss of the ionisable carboxylate group. In addition the chemical shifts of peaks present in the 1H NMR spectrum of the impure naturally occurring 5 matched those of the synthetic analogue (Table S3, ESI†). Interestingly anhydride 6, did not show any signal by ELS (Evaporative Light Scattering) detection and attempts to purify it by HPLC resulted in yields much lower than anticipated. This was attributed to volatility of 5, and was confirmed by HPLC of a sample that had been allowed to evaporate at room temperature. On re-addition of solvent HPLC analysis showed that ca. 96% of the compound had evaporated.

Byssochlamic acid 1 exists in equilibrium with its monohydrated diacid forms which appear together at 9.8 min in the LCMS chromatogram (1a/1b in Fig. 1), as indicated by the characteristic [M + H2O − H]− ion (m/z 349.2) in the negative ESIMS spectrum. This peak appeared in chromatograms of purified byssochlamic acid 1 and of a purified mixture of 1a/1b. When re-injected, this reforms an equilibrium mixture with byssochlamic acid 1. 1H NMR spectra showed that the ring-closed form 1 is preferred in organic solvent, while the ring-open forms 1a/1b are also present in aqueous solvents. Similar behaviour is observed for other maleic anhydride containing metabolites including compounds 5 (see peaks at 7.35 and 4.88 min−1 in Fig. 1) and 6 (Fig. S3, ESI†) and dihydrobyssochlamic acid 2, and for related maleic anhydrides isolated from P. variotii.25

Previous work26 had provided evidence for biosynthesis of byssochlamic acid via dimerisation of maleic anhydride 6 the decarboxylated form of 5. However, it would appear likely that 5 is the actual precursor, decarboxylation providing the exomethylene intermediate 28 necessary for cyclisation with a second molecule of 5 as indicated in Scheme 2 (pathway a). Tautomerisation of the first formed macrocycle leads to byssochlamic acid 1. The heptadrides, agnestadrides A and B can be accounted for by a different mode of dimerisation of these intermediates as indicated in Scheme 2 (pathway b). Formal Michael addition of the anion derived by decarboxylation of 5
now occurs on the other end of the diene system of 28. In the
in vitro studies\(^\text{19}\) leading to 19 strong base was required to trigger
dimerisation. Although presented in Scheme 2 as synchronous
processes, stepwise mechanisms are equally valid at this stage.
Interestingly diene 28 has been isolated as waquafranone B,
along with waquafranone A 29 and several epoxide derivates from
W. aquatica.\(^\text{18}\) Also, the exo-methylene compound 30 has been
isolated, as tubingenic acid, from Aspergillus tubingensis,\(^\text{27}\)
along with the hexyl analogue of 5, itself a metabolite of several
Aspergillus species.\(^\text{26,28,29}\)

Our work with B. fulva has led to the isolation of 5 new maleic
anhydride-containing compounds in addition to byssochlamid acid.
These include dihydrobyssochlamid acid, but more significantly,
two novel 7-membered ring analogues which we have termed
heptadride by analogy to nine-membered nonadrides and eight-
membered octadrides. Heptadrides have never been reported
previously as natural products.\(^\text{‡}\) In addition, the putative mono-
meric maleic anhydride intermediate previously proposed to
undergo macrocyclisation \(\text{via} \) dimerisation to give byssochlamid acid and other nonadrides, has also been isolated for the first
time. A scheme for the biosynthesis of both the nonadrides and
heptadrides is proposed. Work is in progress to establish details
of the biosynthesis, as well as the identification of the biosynthetic
gene cluster.

To account for diversity among nonadride-like natural products,
we propose the generic name \(\text{maleidrides} \) to denote biosynthetically
related compounds, with one (monomaleidrides) or two (bis-
maleidrides) maleic anhydride units anchored on a diversely
substituted ring. Production of the anhydride 5 by \(\text{B. fulva} \),
along with hepta- and nonadrides, indicates that the diversity
among maleidrides very likely arises not only from diversely
substituted anhydride precursors, but also from alternative
dimerisation modes.

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Notes and references

\(\text{‡}\) Tropolone metabolites, e.g. stipitatonic and puberlonic acids, contain
maleic anhydride moieties fused to 7-membered aromatic polyketide-
derived rings, in contrast to the alicyclic rings found in the nonadride
and related metabolites.

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